A Key to the Tadpoles of North Carolina

JOSEPH TRAVIS¹

Department of Zoology, Duke University, Durham, North Carolina 27706

ABSTRACT.— A dichotomous key for identifying the tadpoles of North Carolina and a guide for their field identification with a hand lens are offered. Problems in identifying tadpoles are discussed. The key should be useful throughout the southeastern United States, because it treats most of the species present in this area.

INTRODUCTION

Although anuran larvae are excellent subjects for various types of field and laboratory research, the difficulty of correctly identifying tadpoles is well known. Early keys were either difficult to use (e.g., Wright and Wright 1949) or restricted in taxonomic (e.g., Orton 1952) or geographic (e.g., Smith 1934) coverage. Altig (1970) provided a key to all of the tadpoles found in the continental United States and Canada, and proposed a standard terminology for use in identifying tadpoles. Later references to Altig will be to this 1970 paper. A complete key to variable animals such as anuran larvae can prove difficult to use. Geographic variation can produce ambiguities in identification, and characters that may be diagnostical at a local level may prove impossible to integrate into a more thorough treatment.

North Carolina, with 30 species of anurans (Martof et al. 1980), provides an excellent situation for the development of a more restricted key. The extensive phenotypic variability seen in some species, such as *Hyla crucifer* and *Rana clamitans*, often precludes the effective use of Altig's key in the state. A North Carolina key should also be useful in the southeastern United States, because it includes most of the species found in this area.

My local key was constructed from Altig's and others in the literature, specimens collected by me and others during my four years in North Carolina, and laboratory rearings of unidentified tadpoles. Some key characters used by other workers subsequently proved unreliable and have been deleted. I examined many specimens of 27 of the 30 North Carolina species, including living individuals. *Pseudacris brimleyi, P. brachyphona*, and *Rana heckscheri* were not personally examined. *Hyla versicolor* was obtained from Giles County, Virginia, and specific identity was verified by karyotype analysis. The occurrence of *H. versicolor* in North Carolina remains problematic (Martof et al. 1980).

¹ Present address: Department of Biological Science, Florida State University, Tallahassee, Florida 32306

Joseph Travis

USE OF THE KEY

This contribution should prove useful for live or preserved animals in development stages 25 through 40 (Gosner 1960). The key is not arranged phylogenetically. General arrangement and terminology follow Altig's.

Altig discussed some of the major sources of difficulty in tadpole identification, beginning with the problem of poor preservation. Coloration, useful in identifying live material, will fade in preserved specimens. Labial teeth can fall out, and keratinized skin layers can be dislodged from their underlying structures.

Larval anurans are highly susceptible to environmental influences on morphology. For example, food type can affect mouthpart morphology, causing some distortions of the normal appearance, as is evident in a comparison of laboratory-reared and field-collected tadpoles. Ambient temperatures will affect development rates, and may influence allometric growth patterns (see below). The tails of many tadpoles are damaged by predators, particularly nymphal dragonflies (Caldwell et al. 1980). This may affect comparisons based on length ratios, either because of the tail's abbreviated length or because of a change in its overall shape as regeneration occurs. In addition, a regenerated tail often has a blackened tip or large blotches or large blotches on the fin, marks that may not be part of the normal pattern. Color of a live animal can vary with background. Rana clamitans larvae developing in a pond that contains a high level of gray clay in suspension (as in some ponds in the Sandhills region) will be very pale, while larvae in other situations may range from green to dark brown.

Many characters vary ontogenetically. The most obvious of these is color. The clear belly of young *Rana clamitans* larvae will become an opaque, cream color as the animals develop. The number of rows of labial teeth and the length of the rows change with tadpole age and size. The most subtle ontogenetic variations are the allometric shape changes exhibited by some species. The notable flagellum of a *Hyla femoralis* tadpole is not present in a young larva, but becomes increasingly well developed as the tadpole grows. Many species with broad fins, like *Hyla gratiosa* and *H. chrysoscelis*, have more streamlined profiles as small larvae. All these sources of phenotypic variation should be kept in mind when using any key to tadpoles.

KEY TO THE TADPOLES OF NORTH CAROLINA

1.	Jaws without keratinized sheaths; oral disc and labial teeth absent
	(Microhylidae) Gastrophryne carolinensis
	Jaws with keratinized sheaths; oral disc and labial teeth present
	(Fig. 2) 2
2.	Anus medial (Fig. 4A)
	Anus dextral (Fig. 4B) 4
3.	Oral disc emarginate (Fig. 2); tooth row formula 1-2/3; spiracle distinctly on left
	side of body (Fig. 3A) (Bufonidae) 5
	Oral disc not emarginate (Fig. 2); tooth row formula 2/4 or more; spiracle
	ventrolateral (Pelobatidae) Scaphiopus holbrooki
4.	Oral disc emarginate (Ranidae) 33
	Oral disc not emarginate (Hylidae) 8
5.	P-2 (see Fig. 2) with distinct median gap; P-3 less than 0.50 P-1; papillary
	border extends to lateral tips of P-2;
	light color in life
	P-1 with no median gap; P-3 greater than 0.50 P-1; papillary border extends
	distinctly around P-2; dark color in life
6.	Substantial submarginal papillae (Fig. 2), particularly around emarginate areas
	of oral disc; dorsal tail fin height equal to musculature height (Fig. 1); dorsal fin
	may be higher than ventral fin Bufo terrestris
	Few if any submarginal papillae; dorsal tail fin height lower than musculature
	height; fins subequal in height 7
7.	Dorsum unicolored; snout sloping in lateral view; tail musculature distinctly
	bicolored; tail fin height/musculature height 2.0 or
	less Bufo americanus
	Dorsum often slightly mottled in life; snout rounded in lateral view; tail
	musculature often not distinctly bicolored; tail fin height/musculature height
	greater than 2.0 Bufo woodhousei fowleri
8.	Two rows of posterior labial teeth (Fig. 2)
	Three or more rows of posterial labial teeth 13
9.	A-2 gap (Fig. 2) wide; spiracular tube at least partly free from body wall; body
	slightly depressed; tail tip often solid black (Acris) 10
	A-2 gap narrow to moderate; spiracular tube fully attached to body wall; body
	globular; tail tip, if black, with mottle or blotched black edges 11
10.	Free section of spiracular tube long, almost entire length of tube; throat dark;
	tail musculature finely flecked; Coastal Plain Acris gryllus
	Free section of spiracular tube short, less than or equal to half the length of
	tube; throat light; tail musculature mottled or reticulated; Piedmont and moun-
	tain valleys Acris crepitans
11.	Tail musculature distinctly striped in lateral view; light stripe extends from
	dorsal tail musculature stripe to eye; throat and chest may be mottled; dorsum
	of tail musculature often banded or with
	saddles Limnaoedus ocularis (part)
	Tail musculature not or only faintly striped, but without extension to eye;
	throat and chest light: dorsum of tail musculature not handed 12

Joseph Travis

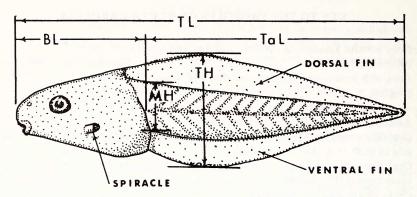


Fig. 1. Left lateral aspect of a tadpole. TL = total length; BL = body length; TaL = tail length; MH = musculature height; TH = tail height. Redrafted from Altig (1970).

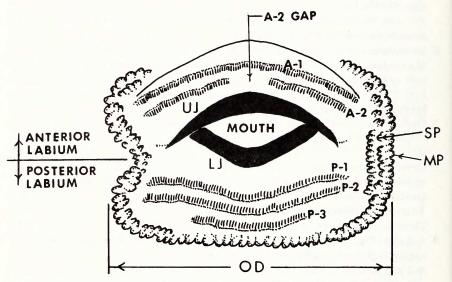


Fig. 2. Tadpole mouth parts, schematic. UJ = upper jaw (mandible); LJ = lower jaw (mandible); A-1,2 = first and second anterior tooth rows; P-1,2,3 = first, second and third posterior tooth rows; SP = submarginal papilla; MP = marginal papilla; OD = oral disc (shown emarginate on viewer's left, marginate on right). Redrafted from Altig (1970).

North Carolina Tadpole Key

13.	Posterior gap in papillary border. 14
1.4	No posterior gap in papillary border
14.	rounded or pointed in dorsal view
	Tail musculature mottled or indistinctly striped, but in no case does a light
	stripe extend forward to eye; snout square in dorsal
	view
15	Tail musculature stripe extends to eye; snout round when viewed dorsally;
	posterior gap in papillary border greater than or equal to length of P-3;
	interocular distance wide, only slightly less than maximal head
	width Limnaoedus ocularis (part)
	No extension of tail musculature stripe to eye; snout tapered or slightly pointed
	in dorsal view; posterior gap in papillary border less than length of P-3;
	interocular distance narrow, substantially less than maximal head
	width Hyla andersoni (part)
16.	P-3 length 0.50 or more times length of P-2; P-3 longer than upper jaw 17
	P-3 length very short, less than 0.50 times length of P-2; P-3 subequal to upper
	jaw
17.	Submarginal papillae absent or few; dorsum of tail musculature usually with
	one black saddle slightly anterior to midlength Hyla gratiosa (part)
	Substantial submarginal papillae; no black saddle on dorsum of tail
10	musculature
18.	Tail musculature distinctly striped; well developed flagellum at tip of tail;
	reddish color in life
	in life
19	Dorsal fin height equal to or greater than musculature height; throat seldom
17.	pigmented; dorsal fin never extends anterior to midway between spiracle and
	eye
	Dorsal fin height less than musculature height; throat pigmented in life; dorsal
	fin extends to posterior border of the
	eye Hyla squirella
20.	Tail musculature striped 21
	Tail musculature not striped 26
21.	A-2 gap wide Pseudacris brimleyi
	A-2 gap narrow
22.	Light dorsal stripe on tail extends to eye; fins clear or with a few stellate
	melanophores; dorsum of tail musculature banded or marked with
	saddles Limnaoedus ocularis (part) Dorsal stripe does not extend to eye; fins clear or mottled; dorsum of tail
	musculature not banded
23.	Tail stripe distinct; snout rounded when viewed dorsally; body slightly de-
_	pressed
	Tail stripe may or may not be distinct; snout squarish or tapering in dorsal view;
	body not depressed 25
24.	Dorsal fin originates anterior to spiracle Pseudacris ornata (part)
	Dorsal fin originates posterior to spiracle Pseudacris triseriata (part)

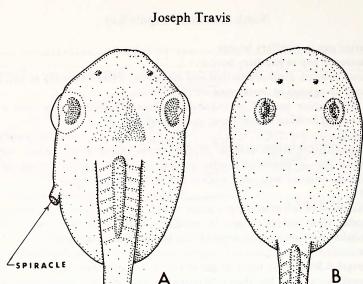


Fig. 3. Eye positions, dorsal aspect. A. Lateral eyes (and spiracle). B. Dorsal eyes. Redrafted from Altig (1970).

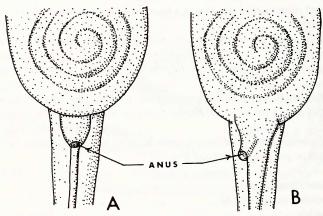


Fig. 4. Anus positions, ventral aspect. A. medial. B. dextral. Redrafted from Altig (1970).

25. Tail stripe faint; snout square when viewed dorsally; snout-spiracle distance/ body length greater than 0.60; spiracle just below eye level; dorsal fin higher than ventral fin; interocular distance only slightly less than maximum head
width Hyla crucifer (part)
Tail stripe distinct; snout tapering in dorsal view; snout-spiracle distance/body length less than 0.60; spiracle well below eye level; fins equal in height;
interocular distance markedly less than maximum head
width Hyla andersoni (part)
26. Total length (Fig. 1) greater than 45
Total length less than 45 mm28

North Carolina Tadpole Key

27.	Jaws wide and rounded; tail musculature unicolored Hyla gratiosa (part) Jaws narrow and angular; tail musculature bicolored
	Pseudacris ornata (part)
28.	Dorsal fin originates anterior to spiracle
20	Dorsal fin originates at or posterior to spiracle
29.	than ventral fin
	Fins and tail musculature mottled or reticulated; fins tapering toward the tail
	tip; dorsal and ventral fins equal in height
30.	Jaws wide and rounded; dorsum of tail musculature usually with black saddle
	slightly anterior to midlength; tail musculature
	unicolored Hyla gratiosa (part)
	Jaws narrow and angled; no black saddle on dorsum of tail musculature; tail
	musculature bicolored Pseudacris ornata (part)
31.	Fins and tail musculature mottled or reticulated; body somewhat globular;
	snout square when viewed dorsally Hyla crucifer (part)
	Fins and tail musculature clear or with a few stellate melanophores; body
	somewhat depressed; snout round when viewed dorsally
32.	Body dark brassy in life; dorsal fin terminates far posterior to
	spiracle Pseudacris brachyphona
	Body color not dark brassy; dorsal fin terminates at or slightly posterior to
22	spiracle
55.	height
	Fins subequal and both lower than musculature
	height
34.	Four or more rows of teeth on anterior or posterior
	labium Rana sylvatica (part)
	Less than four rows of teeth on both anterior and posterior labium 35
35.	A-2 gap ratio greater than 1.5; dorsal fin originates at or only slightly posterior
	to spiracle Rana sylvatica (part)
	A-2 gap ratio variable; dorsal fin originates far posterior to spiracle 36
36.	Lower jaw wide; nostrils medium to large; skin thin, gut visible, with weakly
	pigmented belly in larger specimens; small animals uniform in color, even when
	preserved Rana pipiens group: 37
	Lower jaw narrow; nostrils small; skin thick; gut usually not visible, with
	strongly pigmented belly in larger animals; small animals with gold transverse
	bands on anterior part of body, appearing unevenly pigmented when preserved
37	A-2 gap ratio 2 or more; P-1/P-3 length ratio 1.3 or greater Rana palustris
51.	A-2 gap ratio less than 2; P-1/P-3 length ratio less than 1.5
38.	No keratinized areas at medial tips of P-1; A-2 gap ratio often greater than 1.0;
	color variable Rana sphenocephala
	Keratinized areas present at medial tips of P-1; A-2 gap ratio always less than
-	1.0; color usually dark Rana areolata
39.	Tail musculature unicolored or mottled, but not striped; fins clear or mottled,
	but not in any particular pattern 41
	Tail musculature distinctly bicolored or striped; fins either striped (or with a row of dots) or marked around edges.

Joseph Travis

- 40. Tail musculature distinctly bicolored; fins without stripe; larger specimens have prominent black edging around a clear or speckled fin Rana heckscheri Tail musculature distinctly striped; stripe or row of dots (formed by pigment around the lateral line pores) present on dorsal fin; no black edging on tail

NOTES ON FIELD IDENTIFICATION

Many animals can be diagnosed to genus or species in the field with the use of a hand lens. Small tadpoles are always difficult to identify, but the following notes should allow larger individuals to be placed into one of five principal groups.

- Hylidae: body square in dorsal view, eyes lateral; nostrils small compared to eyes; dextral anus; oral disc not emarginate; never black in color, but can range from bluish to brown.
- Rana: body oval or round in dorsal view, eyes dorsal or dorsolateral; nostrils small compared to eyes; dextral anus; oral disc emarginate; color diagnostically unreliable.
- Bufo: body round or oval in dorsal view, eyes dorsal and with a "crosseyed" aspect; nostrils large, and head appears to have a "snout"; median anus; oral disc emarginate; color may be dark or light (Bufo quercicus).
- Scaphiopus holbrooki: body round or oval in dorsal view, eyes closeset and dorsal; head wide relative to body width; entire body moves from side to side while swimming; median anus; oral disc not emarginate; color black.
- Gastrophryne carolinensis: body round in dorsal view, distinctly depressed; eyes wide-set and lateral; median anus; no oral disc; color dark, although larger individuals have mottled venters and a stripe on the tail musculature.

ACKNOWLEDGMENTS.— Many people helped collect tadpoles, but I owe a particular debt of gratitude to Mr. Jack Longino, Mr. Peter Morin, and Dr. Henry Wilbur. Professor Joseph Bailey allowed me to use the Duke Vertebrate Collection, and Alvin Braswell, North Carolina State Museum of Natural History, graciously provided specimens of preserved tadpoles. Professor Henry Wilbur generously provided me with the time, funding, and encouragement to construct this key and to refine it from 1977 to 1980 while I was at Duke University. Drs. Ronald Altig and John Cooper, and an anonymous reviewer, improved the manuscript considerably through their conscientious suggestions, and Dr. Cooper drafted the figures.

The work that gave rise to this key was supported by NSF grants DEB 76-82620 and DEB 79-11539, both to H. M. Wilbur, and by a postdoctoral fellowship to the author from the University of Virginia through the Mountain Lake Biological Station. Final revisions were made while I was supported by NSF grant DEB 81-82620.

LITERATURE CITED

- Altig, Ronald. 1970. A key to the tadpoles of the continental United States and Canada. Herpetologica 26:180-207.
- Caldwell, Janalee P., J. H. Thorp and T. O. Jervey. 1980. Predator-prey relationships among larval dragonflies, salamanders, and frogs. Oecologia 46:285-289.
- Gosner, Kenneth L. 1960. A simplified table for staging anuran embryos and larvae with notes on identification. Herpetologica 16:183-190.
- Martof, Bernard S., W. M. Palmer, J. R. Bailey and J. R. Harrison. 1980. Amphibians and Reptiles of the Carolinas and Virginia. Univ. North Carolina Press, Chapel Hill. 264 p.
- Orton, Grace L. 1952. Key to the genera of tadpoles in the United States and Canada. Am. Midl. Nat. 47:382-395.

Smith, Hobart M. 1934. The amphibians of Kansas. Am. Midl. Nat. 15:377-528.

Wright, Albert H., and A. A. Wright. 1949. Handbook of Frogs and Toads of the United States and Canada. Comstock Publ. Co., Ithaca, New York. 640 p.

Accepted 16 March 1981